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Title:

Molecular profiling of the metaplastic spindle cell carcinoma of the breast reveals potentially targetable biomarkers

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1 MicroAbstract

- 2 - Spindle cell carcinoma is a rare subtype of metaplastic breast cancer, with triple-negative
3 phenotype. Twenty-three spindle cell carcinomas were comprehensively explored for
4 biomarkers of immuno-oncology and targeted therapies using immunohistochemistry and
5 DNA/RNA sequencing. Spindle cell carcinomas are characterized by targetable
6 molecular alterations in the majority of cases, but due to the lack of uniform findings,
7 individual patient profiling is necessary.

8 Clinical Practice Points

- 9 - The majority of spindle cell carcinomas have triple-negative phenotype.
10 - Its molecular profile is similar to that of other subtypes of metaplastic breast carcinomas.
11 - The molecular alterations within the PIK3CA pathway along with PD-L1 expression
12 characterize a proportion of spindle cell carcinomas and may guide targeted treatments
13 for this rare disease.

14

15

16 **Abstract**

17 **Introduction:** Spindle cell carcinoma is a rare subtype of metaplastic breast cancer (MBC), with
18 triple-negative (TNBC: ER-/PR-/Her2-) phenotype. It is associated with a marked resistance to
19 conventional chemotherapy and has overall poor outcome.

20 **Materials and Methods:** Twenty-three pure spindle cell carcinomas of the breast (18 primary
21 and 5 recurrent/metastatic) were comprehensively explored for biomarkers of immuno-oncology
22 (I-O) and targeted therapies using immunohistochemistry and DNA/RNA sequencing.

23 **Results:** The majority (21/23) of spindle cell carcinomas were TNBC. Estrogen and androgen
24 receptors expression above the therapeutic thresholds were detected in two cases, each.
25 Pathogenic gene mutations were identified in 21/23 cases including *PIK3CA*, *TP53*, *HRAS*, *NF1*,
26 and *PTEN*. One case with matched pre- and post-chemotherapy samples exhibited a consistent
27 mutational profile (*PIK3CA* and *HRAS* mutations) in both samples. Gene amplifications were
28 present in five cases including one case without detectable mutations. The spindle cell
29 carcinomas cohort had consistently low total mutational burden (all below 80th percentile for the
30 entire TNBC cohort). All tumors were microsatellite stable. PD-L1 expression was observed on
31 both tumor cells (TC, in 7/21 cases), and in tumor infiltrating immune cells (IC, 2/21 cases).

32 **Conclusions:** Spindle cell carcinomas are characterized by targetable molecular alterations in the
33 majority of cases, but due to the lack of uniform findings, individual patient profiling is
34 necessary. Detection of individual combinations of biomarkers should improve treatment options
35 for this rare, but aggressive disease.

36 **Key words:** Breast cancer; metaplastic carcinoma; spindle cell carcinoma; molecular profiling;
37 immune checkpoint inhibitors; targeted therapy; mutations

38 Introduction

39 Metaplastic breast carcinoma (MBC) is a rare breast cancer subtype, constituting ~1% of
40 all invasive breast cancers¹. Histologically, MBC is a highly heterogeneous disease,
41 encompassing six different morphologic subtypes including spindle, squamous, chondroid,
42 osseous, rhabdomyoid and mixed morphology¹. Somatic mutations in TP53, PI3K MAPK, RB1
43 and Wnt pathways genes have been frequently described in MBCs²⁻¹¹. MBCs are basal-like and
44 claudin-low breast cancers with a triple-negative phenotype: Estrogen receptor (ER),
45 progesterone receptor (PR) and HER-2/neu negative^{7,9,12-14}. With rare exceptions (low-grade
46 adenosquamous and fibromatosis-like metaplastic variants), MBCs are associated with a high
47 recurrence/metastasis risk, chemotherapy resistance and poor outcome¹⁵.

48 Mutational diversity is reflected in the morphologic heterogeneity of MBCs; *PIK3CA*
49 mutations were detected in all morphologic variants of MBCs, excluding the chondroid variant
50^{5,6,11}, while *TERT* mutations were more prevalent in spindle cell and squamous variants⁵.
51 Microarray expression based studies also revealed differences between the morphologic subtypes
52 of MBC in regards to epithelial-mesenchymal transition (EMT)-related genes such as *CDH1* and
53 *EPCAM*⁷.

54 PD-L1 expression in cancer and/or immune cells, as a predictor of response to immune
55 checkpoint inhibitors, has also been described in a subset of MBCs^{3,9,11,16,17}.

56 Pure spindle cell variants of MBC constitute <10% of all MBCs; the spindle cell pattern
57 is usually seen within a mixed MBC that constitutes ~70% of all MBC morphologies. In the
58 present study, we explored a cohort of pure (>90% of invasive tumor) spindle cell MBC for the
59 biomarkers of response to immuno-oncology (I-O) and targeted therapies.

60

61 **Materials and Methods**

62 **Case selection**

63 Twenty-three pure (>90%) spindle cell MBC identified among cases submitted to Caris
64 Life Sciences (Phoenix, Arizona, USA) for molecular profiling were investigated in the present
65 study. Each case underwent confirmation of the histologic diagnosis, including review of the
66 diagnostic immunohistochemical test results performed at the referring pathology laboratory, by
67 a board-certified pathologist at Caris Life Sciences.

68 Caris Life Sciences de-identified all reports and remnant spindle cell carcinoma samples
69 provided by the referring laboratories. Given that the remnant tissues from previous samplings
70 with no associated identifiers were used, this research was compliant with 45 CFR 46.101(b).
71 Therefore, the present study was deemed exempt from Institutional Review Board approval and
72 consent requirements were waived.

73 **Immunohistochemistry (IHC)**

74 IHC assays included ER, PR, AR, HER-2/neu, PD-L1, and pNTRK. In selected cases,
75 PTEN, cKit and E-cadherin stains were done (the list of antibodies, clones and thresholds for
76 positivity are provided in the Supplemental Table 1).

77 **Next-generation sequencing (NGS)**

78 The samples were profiled using massively parallel sequencing (NGS) of exons from 592
79 genes (SureSelect XT, Agilent, Santa Clara, CA and the NextSeq instrument, Illumina, San
80 Diego, CA)¹⁸.

81 The tumor mutational burden (TMB) was assessed by calculating the number of
82 nonsynonymous missense mutations, excluding common germline variants, in one megabase of

83 DNA. TMB was considered high if ≥ 11 mutations/megabase (mut/Mb) were detected. The
84 estimated threshold was based on a cohort of 603 TNBC cases using an 80th percentile cutoff
85 value as recently suggested by Samstein RM et al.¹⁹. Microsatellite instability (MSI) was
86 calculated from the NGS data by direct analysis of short tandem repeat tracts in the target regions
87 of sequenced genes. The count only included alterations that resulted in increases or decreases in
88 the number of repeats; high microsatellite instability (MSI-H) was defined as ≥ 46 altered
89 microsatellite loci. This threshold was established by comparing NGS with the PCR-based
90 microsatellite fragments analysis results from ~2100 samples^{18,20,21}.

91 Copy number variations (CNVs) were explored by comparing the depth of detected NGS
92 sequence reads to reads from a diploid control. Genes having \geq six copies were considered
93 amplified¹⁸.

94 The ArcherDx FusionPlex Assay (ArcherDX, Boulder, CO) was used for the gene fusion
95 assessment. The gene fusions panel (n=54) is available here:

96 [https://www.carismolecularintelligence.com/wp-content/uploads/2017/03/TN0276-v14_Profile-](https://www.carismolecularintelligence.com/wp-content/uploads/2017/03/TN0276-v14_Profile-Menu.pdf)
97 [Menu.pdf](https://www.carismolecularintelligence.com/wp-content/uploads/2017/03/TN0276-v14_Profile-Menu.pdf).

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105 **Results**

106 **Clinicopathologic characteristics of the cohort**

107 Clinicopathologic data are summarized in Table 1.

108 The study included 23 spindle cell MBCs of which 18 were primary (17 from the breast
109 and one from axilla) and five were recurrent/metastatic cases.

110 All patients were female with a mean age of 60.2 years (range, 30-83 years). With the
111 exception of one case, all were grade 3 carcinomas (Nottingham modification of Bloom-
112 Richardson system), and the majority (21/23) were triple negative. ER and AR (two cases each)
113 expressions above the therapeutic thresholds of 1% and 10% respectively were rarely observed.
114 HER-2/neu was uniformly negative in all cases (0%) (Table 1).

115 **Genomic profile of spindle cell carcinomas**

116 Genomic alterations were detected in 22/23 cases: Twenty-one cases had pathogenic
117 mutations while one case (#11) that was devoid of any detectable pathogenic mutation harbored
118 multiple gene amplifications including *KDR (VEGFR2)*, *KIT*, *PDGFRA*, *FIP1L1*, and *CHIC2*.
119 Only one case (#15) harbored no detectable genomic alterations (Table 1).

120 Mutations most frequently affected *PIK3CA* (10/23, one case was ER+), *TP53* (6/23),
121 *HRAS* and *NFI* (4/23 each), and *PTEN* (3/23) (Supplemental Table 2).

122 Two cases exhibited evidence of epithelial to mesenchymal transition (EMT). The first
123 case (#19, Table 1) was apocrine ductal carcinoma in situ (apocrine DCIS) transitioning into
124 spindle cell carcinoma. Upon separate microdissection analyses, both in-situ and invasive
125 components harbored identical mutational profiles (*PTEN* p.E242fs and *HRAS* p.Q61K
126 mutations). EMT was further evidenced by the loss of E-cadherin and beta-catenin expression in

127 the invasive spindle cell component; however, no mutations were detected in the *CDHI* or
128 *CTNNB1* genes, suggesting possible epigenetic silencing²². AR was positive in an apocrine
129 DCIS, but not an invasive spindle cell component. In the second case (#21, Table 1), a
130 morphologic transition from ductal carcinoma NOS to spindle cell carcinoma was observed. The
131 tumor also harbored a *PTEN* mutation (c.1027-1G>A) and additional *PIK3CA* (p.E542K) and
132 *CDHI* gene mutations (p.E243K, likely pathogenic without E-cadherin protein loss) in both
133 components.

134 One case with available matched pre- and post-chemotherapy samples exhibited a
135 consistent mutational profile (*PIK3CA* and *HRAS* mutations) in both samples. Similarly, another
136 matched case (primary breast and metastatic sample from the lung) had identical mutational
137 profiles at both sites (*PIK3CA* and *KDM6A* mutations).

138 None of the tested spindle cell carcinomas (n=9) exhibited pNTRK positivity by IHC
139 including a case with *NTRK1* gene amplification (Table 1). No *NTRK* gene fusions or any other
140 fusions were detected in any of the successfully tested cases (n=14).

141 Gene amplifications were detected in five of 12 evaluable cases. Two spindle cell
142 carcinomas harbored *CCND1* (encodes cyclin D1 protein) gene amplification. Both cases also
143 had multiple gene amplifications within the fibroblast growth factors family (*FGF3*, *FGF4*,
144 *FGF19* and fibroblast growth factor receptor 3 (*FGFR3*) (Table 1 and Supplemental Table 3).

145 **Immuno-Oncology (I-O) biomarkers in spindle cell carcinomas**

146 The spindle cell carcinomas consistently expressed a low TMB of between 3 and 10
147 muts/Mb. Additionally, all spindle cell carcinomas were microsatellite stable (MSS).

148 One third of the spindle cell carcinomas expressed PD-L1 above the 1% threshold in
149 cancer cells (7/21) (Figure 1, Case#18, upper images); three exhibited diffuse PD-L1 expression
150 in cancer cells (50-100% cancer cell positive, Figure 1A-B). In contrast, PD-L1 expression in
151 immune cells was observed in only two cases, both were triple-negative (Figure 1, case#21,
152 lower images).

153

154 Discussion

155 Recent studies have identified mutations in the TP53, PI3K MAPK, RB1 and Wnt
156 pathways as the most frequent somatic mutations in MBCs²⁻¹¹. Our data confirm that spindle cell
157 MBC shares similar molecular features with other morphologic subtypes of MBCs^{6,9-11,23}.
158 *PIK3CA* mutations are particularly relevant since the ESMO Scale for Clinical Actionability of
159 Molecular Targets (ESCAT) classified them as strong predictors of response to *PIK3CA*
160 inhibitors (level IA) (Supplemental Table 2)^{24,25}. Furthermore, the FDA recently approved the
161 *PIK3CA* inhibitor Piqray (alpelisib) for the treatment of ER-positive and *PIK3CA*-mutated,
162 advanced or metastatic breast cancer following progression on, or after an, endocrine-based
163 regimen. One of the *PIK3CA*-mutated spindle cell carcinomas from our series was ER-positive.
164 In addition, several clinical trials and case studies have revealed promising effects of
165 *PIK3CA*/mTOR inhibitors in patients with advanced/metastatic MBC that harbor mutations in
166 the PI3K pathway^{11,23,26-28}. Basho et al. demonstrated that mTOR inhibitors (temsirolimus or
167 everolimus) combined with doxorubicin and bevacizumab were more effective in the treatment
168 of MBC than in non-MBC²⁸. Similarly, Moulder et al. showed the effectiveness of mTOR
169 inhibitors (temsirolimus) in the treatment of MBC²³. In short, the presence of *PIK3CA*, *PIK3R1*
170 and *PTEN* mutations in ~60% of spindle cell MBC may be a potential therapeutic guide for a
171 substantial proportion of these carcinomas⁶.

172 Mutations in *HRAS* were observed in 17% of the spindle cell MBCs, three of which had a
173 coincident *PIK3CA* mutation. *HRAS* mutations have been well described in other breast cancer
174 subtypes including MBCs^{2,3,10,29,30}. Interestingly, co-occurring *HRAS* and *PIK3CA* mutations
175 have recently been recognized as driver mutations in both benign and malignant
176 adenomyoepitheliomas of the breast^{31,32}. In cell culture models, the *HRAS* p.Q61R mutation

177 appears to drive neoplastic transformation of breast cancer cells followed by reduced E-cadherin
178 expression, increased myoepithelial differentiation and activation of the Akt/PIK3CA pathway.
179 These features, commonly seen in MBC³², underlie the phenotypic similarities between the two
180 entities³³. In our cohort, we clearly demonstrated the EMT in two cases (#19 and 21).

181 Our study also revealed *NF1* gene mutations in a proportion of spindle cell carcinomas.
182 *NF1* germline mutations are responsible for neurofibromatosis type 1 (OMIM#162200) while
183 somatic *NF1* mutations have been described in various cancers including breast cancer^{4,34}.
184 Several previous studies have identified *NF1* mutations in MBC including germline mutations in
185 patients with neurofibromatosis type 1^{4,10,35-40}. Our findings provide further evidence of a role
186 for the *NF1* gene in a subset of MBC.

187 Recently, the FDA approved I-O therapy with atezolizumab for TNBC containing $\geq 1\%$
188 PD-L1 positive immune cells (IC) in the tumor biopsy, based on the IMpassion130 clinical trial
189 (NCT02425891). We found that one third of spindle cell MBC expressed PD-L1; however, it
190 was predominantly expressed in the neoplastic, tumor cell (TC) component. This finding was in
191 line with our previous study of MBC³ and a study by Dill et al.¹⁶. Only two cases in the current
192 study clearly expressed PD-L1 solely in the immune cell (IC) component of the tumor above the
193 companion diagnostics threshold of 1%. For atezolizumab the predictive PD-L1 expression is
194 found in immune cells (in tumors expressing $\geq 1\%$ area occupied by PD-L1+ IC), not in TC
195 expressing PD-L1. This is in contrast to a case study of Adams et al. who revealed an impressive
196 clinical response in a patient with TC PD-L1+ (22c3 clone) advanced MBC treated by combined
197 anti-PD-1 therapy with pembrolizumab and nab-paclitaxel¹⁷. Similarly, Al Sayed et al. reported
198 a complete response to the combination of a novel anti-PD-L1 antibody, durvalumab, with

199 paclitaxel in a patient with chemoresistant, metastatic MBC whose neoplastic cells
200 overexpressed PD-L1⁴¹.

201 In our study, two PD-L1+ (one in TC and IC, respectively) spindle cell carcinomas
202 harbored *PTEN* mutations. *PTEN* mutations in cancer cells may induce immunosuppressive
203 expression signatures and the lack of response to anti-PD-1 therapies⁴². Taken together, PD-L1
204 status in various subgroups of MBC needs to be precisely determined (cell type expressing PD-
205 L1) in the context of additional mutational data (e.g. *PTEN*) and may not unequivocally predict
206 response to I-O therapy. Other, lineage-agnostic predictive biomarkers for immune checkpoint
207 inhibitors (TMB and MSI status) were negative (low TMB and microsatellite stable) in our series
208 of spindle cell carcinomas, similar to the studies of Ng et al.⁶ and Tray et al.⁹. TMB and MSI
209 status in spindle cell carcinomas are also comparable with the data from our large cohort >3000
210 TNBC NOS that exhibited a very low frequency of MSI-H and high TMB⁴³.

211 Determination of the AR status in TNBC is important and positivity has been reported in
212 various subtypes of breast cancer including both TNBC NOS and MBC^{2,44}. Two spindle cell
213 carcinomas from our cohort were also AR-positive. A phase II clinical trial by Gucalp et al.
214 reported AR positivity at 12% among TNBC⁴⁴. A clinical benefit rate was seen in 19% of the
215 patients treated with the anti-AR drug bicalutamide⁴⁴. Another study conducted on 116 TNBC
216 revealed a significant clinical activity of enzalutamide in patients with advanced AR-positive
217 TNBC⁴⁵.

218 Although we found *CCND1* and FGF family genes (*FGF3*, *FGF4*, *FGF19*, and *FGFR1*)
219 amplified in a proportion of spindle cell carcinomas, these genes appear not to be reliable
220 predictors of response to their respective inhibitors in breast cancer²⁴. Therefore, the ESCAT

221 categorized these biomarkers as “Tier X”²⁴ and their clinical relevance in spindle cell carcinomas
222 remains unclear.

223 In conclusion, spindle cell carcinomas are characterized by targetable molecular
224 alterations in the majority of cases, but due to the lack of uniform findings, individual patient
225 profiling is necessary. Detection of individual combinations of biomarkers should improve
226 treatment options for this rare, but aggressive disease.

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228 Conflict of Interest

229 Zoran Gatalica, Phillip Stafford, Jeffrey Swensen, Joanne Xiu and David Spetzler are all
230 employees of Caris Life Sciences. Semir Vranic has received honoraria from Caris Life
231 Sciences. Other authors declare no conflict of interest.

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235 Authors' contributions

236 Conceptualization, Z.G. and S.V.; Formal analysis, Z.G., S.V., P.S., J.P., F.S., J.S. J.X., and
237 D.S.; Writing-original draft preparation Z.G. and S.V.; Writing-review and editing – Z.G. and
238 S.V.; Supervision, Z.G.; Funding acquisition, Z.G. and D.S.

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355 **Tables**

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Case	Site (grade)	TNM Stage (AJCC)	Steroid receptors' status (%)	PD-L1 status (%)	Mutational profile* (NGS)	Copy number variations (NGS)
#1	Primary (3)	Unknown	Negative	Negative	<i>BRAF</i>	None
#2	Primary (3)	Unknown	ER+ (1%)	Negative	<i>TP53</i>	
#3	Primary (3)	pT2NoMx	Negative	Positive (TC+)	<i>PIK3CA, HRAS</i>	
#4	Primary (3)	Unknown	Negative	Negative	<i>KDM6A</i>	
#5	Primary (axilla) (3)	pT3NoMx	AR+ (10%)	Negative	<i>TP53, PIK3CA, NF1</i>	<i>MLLT1</i>
#6	Primary (recurrent) (3)	rpT3NoMx	Negative	Negative	<i>TP53, NF1</i>	
#7	Primary (3)	pT3NoMx	Negative	Negative	<i>NF1</i>	
#8	Primary (3)	pT2NoMx	Negative	Negative	<i>NF1, PIK3R1, BRIP1</i>	
#9	Primary (3)	Unknown	AR+ (15%)	Positive (TC)	<i>TP53, RB1, PTEN</i>	
#10	Primary (recurrent) (3)	Unknown	Negative	n/a	<i>TP53</i>	<i>CYP2D6</i>
#11	Primary (3)	pT3NxMx	Negative	n/a	None	<i>KDR (VEGFR2), KIT**, PDGFRA, FIP1L1, CHIC2</i>
#12	Metastatic (3)	M1	Negative	Positive (TC)	<i>TP53</i>	
#13	Primary (1)	pT3NoMx	ER+ (10%)	Positive (TC)	<i>PIK3CA</i>	<i>FGF4, FGF3, FGF19, CCND1</i>
#14	Primary (postneoadjuvant) (3)	ypT4NoMx	Negative	Positive (TC)	<i>PIK3CA</i>	None
#15	Primary (3)	pT2NoMx	Negative	Negative	None	None
#16	Metastatic (3)	M1	Negative	Negative	<i>KRAS</i>	
#17	Primary (3)	Unknown	Negative	Negative	<i>PIK3CA</i>	
#18	Primary (3)	pT4bNxMx	Negative	Positive (TC)	<i>PIK3CA, HRAS</i>	
#19	Primary (3)	pT2NoMx	Negative	Negative	<i>HRAS, PTEN</i>	None
#20	Primary (postneoadjuvant, matched)*** (3)	ypT1cNoMx	Negative	Negative	<i>PIK3CA, HRAS</i>	<i>AKT2, CCND1, FGF3, FGF4, FGFR3, NTRK1**</i>
#21	Primary (3)	pT2N1aMx	Negative	Positive (IC)	<i>PIK3CA, PTEN, CDH1 E243K</i>	None
#22	Primary (3)	Unknown	Negative	Positive (IC)	<i>PIK3CA E545K; NF2 V219fs</i>	None
#23	Primary and meta (matched) (3)	M1	Negative	Positive (100% TC)	<i>PIK3CA Q546K, KDM6A E138I</i>	None

357 *Only pathogenic mutations are listed.

358 ** Both cases were further tested by immunohistochemistry (CD117 and panTRK antibodies) and were negative.

359 ***Matched core and surgical biopsy were tested; this cancer was treated with neoadjuvant chemotherapy but the
360 tumor was chemoresistant.

361 n/a = Not available

362 TC = Tumor cells; IC = Immune cells

363 ER = Estrogen receptor; PR = Progesterone receptor; AR = Androgen receptor

364 NGS = Next-generation sequencing

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366 **Table 1.** Molecular profiling features of the spindle cell carcinoma cohort.

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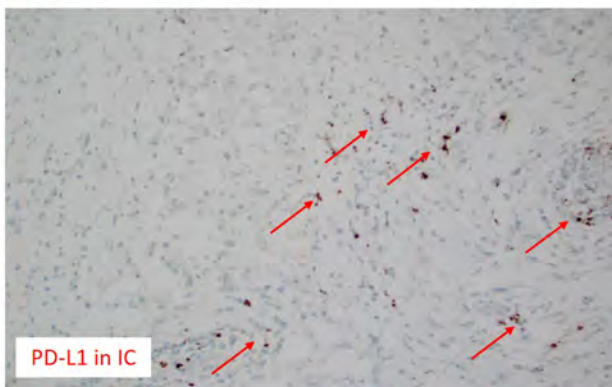
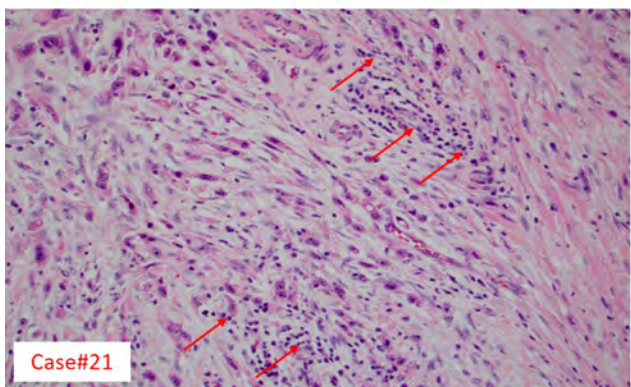
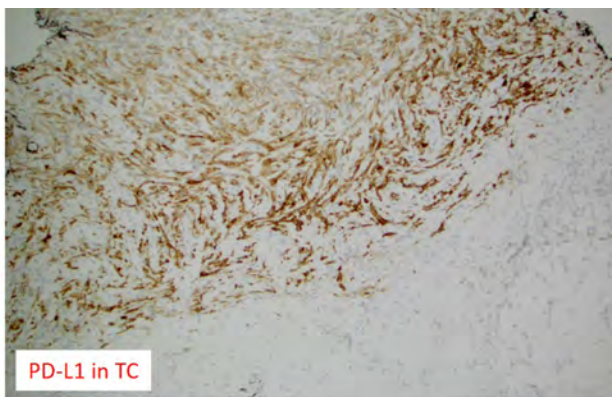
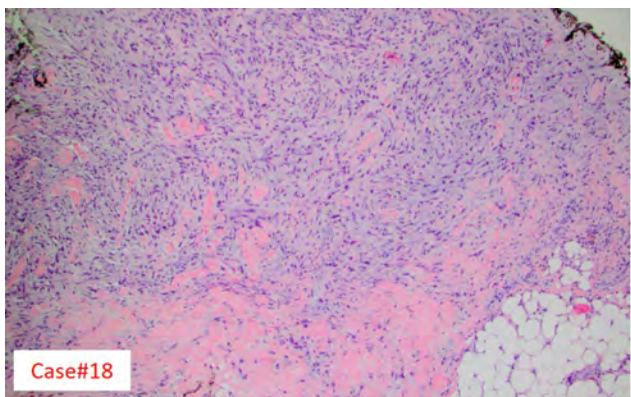
371 **Figures**

372 **Figure 1.** Two triple-negative spindle cell carcinomas with PD-L1 positivity: Case#18 (upper
373 two figures) with diffuse (70%) PD-L1 expression in cancer cells (TC); Case#21 (lower two
374 figures) showing PD-L1 positivity at 1% in immune cells (red arrows). The left-sided images
375 represent hematoxylin-eosin (H&E) stained slides; both cases were tested with VENTANA PD-
376 L1 (SP142) Assay, FDA-approved test.

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