Review

SARS-CoV-2 Infection and Smoking: What is the association? A brief review

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## SARS-CoV-2 Infection and Smoking: What is the association? A brief review

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#### **Abstract**

Susceptibility to severe illness from COVID-19 is anticipated to be associated with cigarette smoking as it aggravates the risk of cardiovascular and respiratory illness, including infections. This is particularly important with the advent of a new strain of coronaviruses, the severe acute respiratory syndrome coronavirus (SARS-CoV-2) that has led to the present pandemic, coronavirus disease 2019 (COVID-19). Although, the effects of smoking on COVID-19 are less described and controversial, we presume a link between smoking and COVID-19. Smoking has been shown to enhance the expression of the angiotensin-converting enzyme-2 (ACE-2) and transmembrane serine protease 2 (TMPRSS2) key entry genes utilized by SARS-CoV-2 to infect cells and induce a 'cytokine storm', which further increases the severity of COVID-19 clinical course. Nevertheless, the impact of smoking on ACE-2 and TMPRSS2 receptors expression remains paradoxical. Thus, further research is necessary to unravel the association between smoking and COVID-19 and to pursue the development of potential novel therapies that are able to constrain the morbidity and mortality provoked by this infectious disease. Herein we present a brief overview of the current knowledge on the correlation between smoking and the expression of SARS-CoV-2 key entry genes, clinical manifestations, and disease progression.

Keywords: SARS-CoV-2; Coronavirus; COVID-19; Smoking; Lung Disease; Oral Disease

#### **List of Abbreviations**

ACE2: Angiotensin-converting enzyme-2; ACEIs: Angiotensin-converting enzyme inhibitors; ADAM17: ADAM metallopeptidase domain 17; ALCAM: Activated leukocyte cell adhesion molecule; Ang: Angiotensin; ARBs: Angiotensin receptor blockers; ARDS: Acute respiratory distress syndrome; BatCoV: Bat coronavirus; CLDN7: Claudin 7; COPD: Chronic obstructive pulmonary disease; COVID-19: Coronavirus disease 2019; CTNNB1: Catenin beta 1; ERK: Extracellular signal-regulated kinases; HDAC6: Histone deacetylase 6; HIV-1: Human

immunodeficiency virus 1; hrsACE2: human recombinant soluble ACE-2; IFN: Interferons; IPF: Idiopathic pulmonary fibrosis; IR: Ionizing radiation; JNK: c-Jun N-terminal kinase; MERS: Middle-East respiratory syndrome; MCN: Mucin; R0: R-nought; nAChR: α7 nicotinic acetylcholine receptor; NO: Nitric oxide; RAS: Renin-angiotensin; RR: Relative risk; SARS-CoV-2: Severe acute respiratory syndrome coronavirus; TJP3: Tight junction protein 3; TMPRSS: Transmembrane serine protease

### Introduction

The current outbreak of a novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) causes a severe respiratory illness called coronavirus disease 2019 (COVID-19). SARS-CoV-2 is an enveloped RNA virus with high zoonotic potential. Genetic analysis revealed that SARS-CoV-2 belongs to the *Betacoronavirus* genus and the *Sarbecovirus* subgenus, indicating that it originated from the bat coronavirus (BatCoV RaTG13) [1]. The initial epidemic, which began in Wuhan, China, continued seeding secondary chains through international travel and propagated to over 180 countries and territories [2]. As of October 2020, more than 34 million confirmed cases have been reported, including over one million deaths, with a rising number of cases reported in India, the United States, and Brazil [3]. SARS-CoV-2 is an extremely contagious infection with interpersonal transmission by aerosols and fomites; the virus remains environmentally active in the atmosphere and on surfaces for several days [4]. These features make it possible for the virus to spread rapidly and cause a global pandemic.

The outbreak was initially presented in the form of pneumonia of unknown etiology. As the epidemic evolved into a global pandemic, it became clear that the SARS-CoV-2 infection contributes to multi-organ disease, with a high incidence of acute respiratory distress syndrome (ARDS) and digestive tract complications [5]. SARS-CoV-2 has a broad spectrum of clinical manifestations ranging from asymptomatic to multi-organ dysfunction [6]. The disease is graded into mild, moderate, severe, and critical, depending on the severity of the presentation, while the most common symptoms include fever, cough, diarrhea, and fatigue [6]. Based on the R-nought (R0) values, it is considered that SARS-CoV-2 is more contagious than previous coronavirus pandemics (SARS or MERS) [7, 8]. Certain population segments are at a higher risk of fatality, including the elderly, immunocompromised, comorbid patients, and smokers [9,

10]. In general, smokers are more likely to get respiratory diseases than non-smokers [11]. Smokers also have twice the risk of acquiring influenza infection, tuberculosis, and bacterial pneumonia [12, 13]. A meta-analysis revealed that 22% (n = 31/139) of current smokers and 46% (n = 13/28) of ex-smokers had severe complications, including ARDS [14]. However, relative risk (RR) analysis showed that current smokers were 1.45 times more likely than former and never-smoker patients to have severe complications [14-16]. Nevertheless, the role of smoking in COVID-19 remains controversial [14, 17, 18]. This review will focus on the relationship between smoking and receptors of SARS-CoV-2 and impact on COVID-19 leading to tissue damage and diseases especially in the lungs.

### SARS-CoV-2 and its key entry genes

The Latin word "corona" ascended from the crown-like appearance of the coronavirus structural images. The large type 1 transmembrane spike (S) glycoproteins attribute to the virus's corona shape. The S glycoprotein comprises two distinct functional domains (S1 and S2) that are assumed to mediate host cell binding, membrane fusion, and virus entry [19]. The S1 domain contains receptor-binding sites for *angiotensin-converting enzyme-2 (ACE2)* and is responsible for virus entry into host cells [20]. The S2 domain enables the fusion of the virus membrane and the host cell, essential for cell infiltration [21].

There is still a limited understanding of the pathogenic pathways of SARS-CoV-2. Viral particles infiltrate the host respiratory epithelial cell through metallopeptidase cell identification. Host cell enzymes must cleave the S glycoprotein at two different sites for coronaviruses to enter target cells, thereby presenting potential drug targets. Several host enzymes are known to activate SARS-CoV-2, including cathepsin L, furin, and transmembrane serine protease 2 (TMPRSS2), TMPRSS4, TMPRSS11A, and TMPRSS11D [22]. However, *TMPRSS2* and *furin* play key roles in the proteolytic activation of SARS-CoV-2 [23]. Furin and TMPRSS2 are essential enzymes for S protein's cleavage at the S1/S2 and the S2' sites, respectively [23]. Bestle et al. [23] have shown that *TMPRSS2* is necessary for SARS-CoV-2 S glycoprotein priming through *in-vitro TMPRSS2* knockdown in human epithelial cells. Similarly, they revealed that synthetic *furin* inhibitors strongly blocked the transmissibility of

SARS-CoV-2 in lung epithelial cells and that more potent antiviral effects were obtained by combining *TMPRSS2* and *furin* inhibitors [23].

SARS-CoV-2 host moiety, the *ACE2* receptors, are essential for marinating human's body homeostasis, including membrane trafficking, the renin-angiotensin (RAS), and cardiovascular systems [24-26]. *ACE2* is widely expressed in all tissues with relatively higher expression in the respiratory system, including type I and type II alveolar cells, the central nervous system, the cardiovascular system, kidneys, and the gastrointestinal tract [27]. *ACE2* acts as a key regulator of RAS, primarily by transforming Ang (angiotensin) I to Ang 1–9 and Ang II to Ang 1–7 [28]. *ACE2* activity occurs in various lung diseases, including lung injury and fibrosis, pulmonary hypertension, and ARDS [29]. More recently, the role of *ACE2* in COVID-19 pathogenesis has gained considerable attention due to its critical link with immunity, inflammation, the digestive tract, and cardiovascular diseases. SARS-CoV-2 binding affinity for *ACE2* correlates with the viral replication rate, transmissibility, and disease severity [30]. *ACE2* and viral S protein interactions are considered a promising therapeutic target for vaccine production [31].

We performed a comprehensive literature search using the PubMed/MEDLINE/ database to explore papers published until 31 January 2021. We used the following keywords: COVID-19, SARS-CoV-2, smoking, *ACE2*, *TMPRSS2*, *furin*, and treatment. Retrieved publications were selected independently for their relevance and contribution; literature search was constrained by study subjects (in-vitro, in-vivo, humans) and language (English). Furthermore, references retrieved from articles and those from recent reviews of smoking and COVID-19 were analyzed. This review also includes publications focusing on epidemiology and clinical characteristics of COVID-19, the association of smoking with COVID-19, in addition to the management and therapeutic options for COVID-19.

### Smoking as a risk factor for COVID-19

Smoking is an established risk factor for several cardio-metabolic and respiratory diseases, including chronic obstructive pulmonary disease (COPD) and bronchial asthma [32]. Smoking tobacco promotes exposure to several toxic chemicals, including 1,3-butadiene, benzene, and

NO<sub>2</sub> [33]. It can induce inflammation of the respiratory tract, allergy, permeability of epithelial cells, mucus formation, and impair mucociliary clearance [34].

Robust evidence indicates that the risk of *Mycobacterium tuberculosis* infection is almost doubled by smoking-induced immune suppression. Likewise, the risk of influenza, *mycoplasma pneumoniae*, *legionella*, and *pneumococcal* infections is 3-5 times higher in smokers. [35]. However, literature on the role of smoking in the pathogenesis of previous coronavirus outbreaks (Middle Eastern respiratory syndrome coronavirus (MERS) and severe respiratory syndrome coronavirus (SARS)) is scarce. One study from the Republic of Korea, based only on twenty-six samples, showed that smoking increased the risk of death in MERS patients [36]. Although data is sparse for COVID-19 cases, we have summarized available data comparing disease fatality in smokers and non-smokers (Table 1).

**Table 1: Smoking and COVID-19** 

City/Country	Reported cases (%)	Smoking history		Co- morbidity (%)	Mortality (%)	Reference
		Severe cases n (%)	Mild cases n (%)			
Wuhan/China	140 (100%)	2/58 (3.4%)	0/82 (0%)	90 (64.3%)		[37]
Sindh/Pakistan	382 (41.5%)	Positive correlation with disease severity and death		Positive correlation	2.8%	[38]
China	1590 (100%)	111/1590 (7%)		399 (25%)		[39]

Wuhan/China	191 (100%)	5/54 (9%) of dead	6/137 (4%) of survivors		54 (28.2%)	[40]
Wuhan/China	78 (100%)	3/11 (27.3%)	2/67 (3%)	29 (37.1%)	(2.5%)	[41]
Chongqing/China	133 (100%)	58/65 (89.71%)	61/68 (89.23%)	133 (100%)	75	[42]
China	1099 (100%)	29/172 (16.9%)	108/913 (11.8%)	261 (23.7%)	(1.4%)	[43]
Wuhan/China	41 (100%)	0/13 (0%)	3/28 (11%)	(32%)	6 (15%)	[44]

Several COVID-19 cohort studies analyzed the impact of smoking on disease incidence, comorbidity, and mortality, the results are summarized in Table 1. The studies either reported a correlation between smoking and COVID-19 severity or divided cohorts into mild and severe cases, with percentages of smokers in each group. According to Ujjan et al. [38], there is a correlation between smoking and disease severity, with all deceased patients (2.8%) being smokers. Similarly, Zhou et al [40] studied 191 inpatients, 54 of whom died while in the hospital and 137 of whom were discharged. They found that dead patients had a greater smoking tendency than recovering ones. Although several studies found a link between disease severity and smoking [41, 45], few found no difference or a negative correlation between smoking and COVID-19 severity [42, 44].

Zhao et al. (2020) conducted a meta-analysis of seven studies analyzing the association of COVID-19 fatality with smoking. The authors found that smoking doubled the risk of severe COVID-19 [46]. Similarly, a meta-analysis of 13 published Chinese studies found that age over 65, male gender, and smoking were risk factors for disease progression in COVID-19 patients [47]. Another meta-analysis conducted by Patanavanich and Glantz (2020) showed a significant

association between smoking and the progression of COVID-19 [48]. Moreover, another study showed that being a smoker or former smoker was a greater risk factor for a severe COVID-19 infection (OR = 1.96, CI = 1.36 - 2.83) and posed a greater likelihood of a more critical condition (OR = 1.79, CI = 1.19 - 2.70) [32].

However, on the contrary, studies have demonstrated a lack of association between smoking, COVID-19, and the severity of COVID-19 disease. In contrast, studies carried out in China reported only 25% of the COVID-19 patients to be smokers [15, 39, 49-51], in Italy, as low as 15% of COVID-19 patients were former smokers [52].

### Impact of Smoking on the expression of SARC-CoV-2 key entry genes

Chronic smokers or prolonged exposure to smoke tends to display several co-morbidities, including emphysema, atherosclerosis, and immune dysregulation [53], which further enhances COVID-19 progression.

Hung et al. [54] found that exposure to smoke increased the pulmonary activity of *ACE2* and that *ACE2* knockout mice had significant pulmonary inflammation and distress in response to cigarette smoke. Smoking increased pulmonary JNK, p38, and ERK1/2 levels, indicating that *ACE2* expression caused by smoking promotes inflammation and lung injury [54]. Similarly, smoking-induced abnormal expression of the ACE2 pathway is associated with blood gas changes, lung inflammation, edema, and injury [55]. The nitrogen dioxide present in cigarette smoke enhances the binding of *ACE2* to its receptor up to 100 times by stimulating ACE enzyme activity [33]. Radzikowska et al. (2020) reported that smoking, asthma, obesity, and hypertension could elevate *ACE2* expression in the bronchial biopsy, bronchioalveolar lavage, or blood samples [56]. Similarly, Brake et al. (2020) reported that smoking upregulates ACE2 receptor expression in COVID-19 patients [57]. Further, studies have found elevated expression of *ACE2* and *TMPRSS2* in several groups of patients with chronic obstructive pulmonary disease (COPD) and idiopathic pulmonary fibrosis (IPF) [53, 58-62]; both expressions significantly correlated with previous cigarette exposure [58, 63, 64]; COPD is considered a major risk factor for COVID-19 [65, 66].

However, *ACE2* expression was not altered in patients with asthma or pulmonary sarcoidosis [53, 67, 68]. On the contrary, Matusiak and Schürch [69] conducted a study in asthmatic patients and found loss of *ACE2* expression in nasal epithelium and increased *TMPRSS2* expression in bronchi and central airways. Similarly, Liu et al. (2020) recorded that plasma *ACE2* level of COVID-19 patients was significantly elevated and correlated linearly with viral load and lung injury [70]. Another study by Leung et al. (2020) analyzed *ACE2* expression in the small airway epithelia of COPD patients and indicated that smoking frequency positively correlates with *ACE2* gene expression, which was higher in current smokers compared to never-smokers [71]. Since *ACE2* is considered an interferon-stimulated gene [72], corticosteroids can be attributed to one of the possible reasons for the loss of *ACE2* expression in patients with asthma. However, no significant difference was found in furin expression in both asthmatics and healthy individuals [69]. Furthermore, *in-vivo* studies also showed unaltered *ACE2* expression in a mouse model of cystic fibrosis as well as in mice exposed to several carcinogens (arsenic, ionizing radiation (IR), and 1,3-butadiene) [53, 73-76].

Halwani et al. [58] used public gene expression datasets to compare the expression of *ACE2* and *TMPRSS2* in blood samples from children and adults and found no significant difference between them. There was also no difference in the expression of *ACE2* and *TMPRSS2* in COPD and diabetic patients, while there was an increase in both enzymes' expression in hypertensive patients [58]. However, in asthmatic patients, only *ACE2* expression was elevated [58]. Additionally, the expression of *ACE2* and *TMPRSS2* was lower in children's airways than smokers and patients with COPD, suggesting a plausible reason for the difference in the severity of COVID-19 disease among different groups of patients [58].

A recent study by Cai et al. [77] analyzed transcriptomic data sets regarding the association between smoking and the expression of SARS-CoV-2 receptors, *ACE2*, *TMPRSS2*, and *furin* in lung tissues. They found an upregulation of *ACE2* expression in smokers compared to non-smokers with a 25% increase in lung tissue expression [77]. Another study analyzed *ACE2* in both mice and human lung tissues and found enhanced expression of *ACE2* in smokers than non-smokers [53]. Cigarette smoke exposure increased *ACE2* expression in mice by 80 percent. Similarly, exposure to smoke in human lung epithelial cells showed increased *ACE2* expression

by 30-55% relative to non-smokers [53]. Smith et al. [53] identified ACE2 as an interferonstimulated gene in the lung epithelium's secretory cells. They indicated that SARS-CoV-2 infections could produce positive feedback loops, further inducing ACE2 levels and allowing viral dissemination. Moreover, Cai et al. [77] examined the effects of smoking on the pulmonary expression of ACE2 in single bronchial epithelial cells. They found that smokinginduced morphological changes in cells of the bronchial epithelium. ACE2 was also detected in secretory goblet cells in the airway of cigarette smokers [53, 77], whereas in non-smokers ACE2 was only expressed in alveolar type II cells [77, 78], suggesting elevated ACE2 expression in smokers' lungs as a plausible derivate of smoking-induced secretory cell hyperplasia. Furthermore, single-cell sequencing data showed that ACE2 levels were significantly associated with multiple mucin genes (MUC1, MUC4, MUC15 and MUC16) and other epithelial barrier associated genes (ALCAM, [79] CLDN7, [80] and TJP3 [81]) [53]. Also, gene ontology analysis showed that ACE2-associated transcripts correlate with genes regulating secretion, glycosylation and respond to toxic substances [82]. Besides, interferons induced significant ACE2 expression. Although IFN-α, IFN-β, and IFN-γ treatment-induced ACE2 expression in tracheal cells, only IFN- $\alpha$  and IFN- $\beta$  altered ACE2 expression in small airway cells [53, 83, 84], suggesting that ACE2 expression can be plausibly induced by either viral infections or interferon treatment [53]. In comparison to non-smokers, the main facilitators of ACE2 receptor in smokers are ADAM17 [85], androgen receptor (AR) gene [86].

Another study looked at the relationship between *ACE2* expression and various smoking methods and reported that smoking but not vaping increases *ACE2* expression [87]. Furthermore, the study found that smoking nicotine and flavored e-cigarettes led to increased pro-inflammatory cytokine and inflammasome production [87]. Similarly, nicotine upregulates *ACE2* expression [88] by the activation of α7 nicotinic acetylcholine receptor (nAChR) on bronchial epithelial cells [89, 90], suggesting that smokers are at a higher risk for SARS-CoV-2 infection. Moreover, nitric oxide (NO) is also present in cigarette smoke and may impair immunity, raising the risk of SARS-CoV-2 infection [91]. Although studies have indicated NO as a risk factor for SARS-CoV-2 infection, the role of nicotine and NO in COVID-19 is controversial. In contrast, various studies indicated controversial data regarding the role of cigarette smoke regulation of *ACE2* expression. An in-vitro study analyzed cigarette smoke's

effect on ACE2 expression using human bronchial epithelial cells (H292) by an air-liquid interface system [92]. The study demonstrated that cigarette smoke exposure induced loss of ACE2 mRNA expression and did not find any association between ACE2 expression and release of IL-6 from H292 cells exposed to smoke; thus, suggesting that loss of ACE2 in lung cells can be either under the direct effect of nicotine or, some other constituent in cigarette smoke [92]. Moreover, this study indicated that ACE2 mRNA levels correlates both in smokers and non-smokers and cigarette smoking acts selectively on the bronchial epithelium by inhibiting viral infection and stimulating other genes' expression that can activate several transcriptomic pathways [92]. In addition to mRNA levels, the study analyzed protein expression of ACE2 and reported inconsistency between mRNA and protein levels of ACE2; indicating the protein expression profile is plausibly related to the fact that enhanced mRNA expression can be a consequence of compensatory mechanism for the loss of ACE2 protein expression on the cellular membrane [92]. The study also suggests a plausible pharmaceutical role of nicotine in COVID-19 treatment [92]. Few studies showed that nicotine has antiinflammatory activity by reducing inflammation via nAChR α7 subunit on macrophages [17, 51], thus, possibly adding a therapeutic value. Similarly, NO was shown to display beneficial effects as a pulmonary vasodilator [93] and reduced the risk of severe COVID-19 disease in smokers [94].

A previous study demonstrated higher *ACE2* activity in male mouse kidneys and adipose tissues than female mice; however, the study did not report high *ACE2* expression [95]. Similarly, another study also reported higher renal *ACE2* activity in males compared to females; lower *ACE2* activity in female kidneys was found to be due to the presence of E2 in the ovarian hormone [96]. However, the study reported no sex differences in *ACE2* activity in the heart and lung [96, 97]. Moreover, research has demonstrated that sex hormones (androgens and estrogens) regulate the renin-angiotensin system [98-101]; while androgens increase plasma renin activity [98], estrogens reduce plasma renin activity [99]. Based on this, Majdic (2020) proposed that sex hormones can modulate ACE2 expression in the lung. Consequently, this could be one of the underlying factors for gender disparities in COVID-19 morbidity and mortality [102]. As published, the COVID-19 death rate was found to be dependent on sex; in China [103] and Italy [104], men's death rate was significantly higher than in women. ACE2

expression was significantly lower in female primary isolated human airway smooth muscle cells than males using western blot analysis [105]. Moreover, the study demonstrated that exposure to testosterone and estrogen significantly upregulated and downregulated *ACE2* expression, respectively [105].

Moreover, sex hormonal regulation was found to regulate ACE2 and TMPRSS2, resulting in differential gender susceptibility to COVID-19. While ACE2 expression increased in females either due to skewed chromosome X inactivation or by estrogens, reduced androgen levels in women resulted in low TMPRSS2 expression increasing its protective role against COVID-19 development and progression [106]. This indicates that the role of sex hormones and chromosomes influence the discrepancy in the severity of COVID-19 infection between sexes, thus signifying the increased susceptibility to COVID-19 in men [106]. Recently, Chakladar and colleagues [86] found co-upregulation of both the SARS-CoV-2 receptors, ACE2 and TMPRSS2, in smokers compared with non-smokers. The study used gene set enrichment analysis (GSEA) and found the androgen signaling pathway pertinent to ACE2 and TMPRSS2 expression [86]. Voinsky and Gurwitz [107] reported elevated TMPRSS2 expression in bronchial samples from smokers compared with non-smokers. Studies have shown that smoking enhances the androgen hormone levels [108], which increases the expression of ACE2 and TMPRSS2 receptors [86, 109]. Along with the stimulation of the androgen signaling pathway, upregulation of the receptors also correlated with the central regulators' overexpression (HDAC6, CTNNB1, and SMARCA4) of the androgen pathways [86]. Increased TPMRSS4 expression in lung epithelial cells of smokers may be due to prolonged exposure to several compounds in tobacco smoke, including nicotine [110], acetaldehyde [111], and tar [112], resulting in oxidative stress and bronchial inflammation [113-115]. On the contrary, Cai et al. [77] did not find any correlation between smoking and TMPRSS2 expression. Similarly, another study found that smoke-exposure enhanced Cathepsin B, but not TMPRSS2 or Cathepsin L expression in the respiratory tract of mice and humans [53]. Moreover, analogous to lung epithelium, in smokers, ACE2 and TMPRSS2 expression was enhanced in the oral epithelium; thus, suggesting a high susceptibility of SARS-CoV-2 in oral epithelial cells [86].

Based on the studies reported, it is controversial whether smoking increases or reduces the risk of contracting COVID-19. A study in Atlanta performed analysis across six acute care hospitals and associated outpatient clinics in 220 hospitalized and 311 non-hospitalized COVID-19 patients and reported smoking was an independent risk factor for COVID-19 hospitalization [116]. Moreover, meta-analysis reported smokers to be at a higher risk of developing severe or progressive disease [117]. Unlike smoking, there is no data to support vaping prevalence in COVID-19 patients; nevertheless, it has been hypothesized that vaping could prime the lung for SARS-CoV-2 infection and correlate with worst outcomes, chiefly based on ancillary data on lung inflammatory processes described in in-vitro and in-vivo studies [118, 119]. However, epidemiological studies failed to report vaping habits in comparison to reports of smoking amongst hospitalized COVID-19 patients [120]. While, a study from the University of Stanford, based on a self-reported internet survey, revealed that use of e-cigarette increases the risk by five-fold of positive COVID-19 in comparison to never users [121], on the contrary, cross sectional self-reported surveys demonstrated lack of association between diagnosed/suspected COVID-19 among never, current, and ex-vapers [122, 123]. In addition, Farsalinos and colleagues reported that current smoking status was comparatively lower in COVID-19 patients; thus, indicating a protective role of smoking against COVID-19 [17]. Although smoking may not plausibly enhance risk for developing COVID-19, the biological and inflammatory signaling cascade that occurs during SARS-CoV-2 infection can be potentially devastating for a smoker.

### **Possible Therapeutic Targets**

Drugs including chloroquine and hydroxychloroquine are currently approved for malaria and autoimmune diseases. Several countries have widely used them for COVID-19 treatment [124, 125]. However, due to their adverse side-effects, including a loss of vision, nausea, and cardiovascular complications, neither chloroquine nor hydroxychloroquine have been recommended in the clinical setting to treat COVID-19. It is shown that chloroquine and hydroxychloroquine exert their effect by accumulating in lymphocytes and macrophages, thus inhibiting pro-inflammatory cytokines secretion and activating anti-SARS-CoV-2 CD8+T-cells [126]. Several other immunomodulators and anti-inflammatory drugs are also being tested in clinical trials [127].

On the other hand, novel therapeutic interventions are urgently needed to stop or slow down the virus's replication and spread. Many therapeutic options are currently being investigated apart from vaccine development, including drugs with distinct antiviral and immune-inflammatory properties [40, 128]. The efforts are directed to mask the COVID-19 functional receptors on human cells, *ACE2* and *TMPRSS2* [40, 128]. Several therapeutic strategies for COVID-19 have been recommended, including pre-existing medicines (chloroquine, hydroxychloroquine, remdesivir and lopinavir) for viral treatment, protein modeling, and genetic analysis using several computing techniques [129-133]. Camostat, the serine protease inhibitor, blocks both *TMPRSS2* and *TMPRSS4* and reduces influenza virus replication and cytokine production in primary cultures of human tracheal epithelial cells [134]. Therefore, camostat was proposed as a potential COVID-19 therapeutic agent [135].

Reducing or quitting smoking can slow down the damage of the respiratory epithelial architecture [136], reduce secretory cell hyperplasia [137], and decrease *ACE2* secretion by 40% [53]. Soluble ACE2 is considered a plausible treatment option in cases where *ACE2* is over-expressed in the airway's epithelium in smokers [138]. Monteil et al. [139] showed that human recombinant soluble ACE-2 (hrsACE2) reduced SARS-CoV-2 viral loads in infected Vero-E6 cells, kidneys, and vascular organoids. In Europe, hrsACE2 is under Phase 2 clinical trial (ClinicalTrials.gov NCTO4335136) as a therapeutic agent for COVID-19 [140]. However, the efficacy of such therapy in smokers and COPD patients who have elevated levels of *ACE2* needs to be determined. Moreover, use of therapeutics and dietary supplements to normalize NO levels in smokers should be considered to minimize the incidence of COVID-19 disease and its complications [91].

Investigations using *in-vitro* models and anti-retroviral drugs (lopinavir and ritonavir) are effective against human immunodeficiency virus 1 (HIV-1) infection, SARS-CoV, MERS-CoV, and SARS-CoV-2 viruses [141, 142]. However, research has shown that neither lopinavir nor ritonavir improves symptoms compared to standard care in severe COVID-19 patients [143, 144]. However, a combination of lopinavir-ritonavir with oseltamivir showed complete

recovery after COVID-19 pneumonia [145, 146]. On the other hand, remdesivir has been shown to effectively inhibit SARS-CoV-2 in *in-vitro* studies using Vero E6 cells [129, 147].

## **Summary and Outlook**

Although smoking can enhance the expression of key entry genes of SARS-CoV-2 used for viral activation, the underlying mechanisms of tobacco-related upregulation of these receptors and the degree to which smoking affects infection susceptibility and clinical manifestations remain unknown. Future mechanistic studies are warranted to address these issues.

#### **Declarations**

#### **Author Contributions**

"Conceptualization, A.E.A.; writing—original draft preparation, I.G. and M.U.S.; writing—review and editing, S.V., M.A.A., K.E.E., A.H.A. and A.E.A. All authors have read and agreed to the published version of the manuscript."

# Both authors contributed equally to this manuscript

## **Declaration of Competing Interest**

"The authors declare no conflict of interest." "The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results".

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#### References

- 1. Zhou, P., et al., *A pneumonia outbreak associated with a new coronavirus of probable bat origin.* 2020. **579**(7798): p. 270-273.
- 2. Organization, W.H., Coronavirus disease (COVID-19): weekly epidemiological update. 2020.
- 3. Dong, E., H. Du, and L.J.T.L.i.d. Gardner, *An interactive web-based dashboard to track COVID-19 in real time*. 2020. **20**(5): p. 533-534.
- 4. Van Doremalen, N., et al., *Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1*. 2020. **382**(16): p. 1564-1567.
- 5. Cascella, M., et al., Features, evaluation and treatment coronavirus (COVID-19), in Statpearls [internet]. 2020, StatPearls Publishing.
- 6. Wang, Y., et al., *Unique epidemiological and clinical features of the emerging 2019 novel coronavirus pneumonia (COVID-19) implicate special control measures.* 2020. **92**(6): p. 568-576.
- 7. Ramadan, N. and H. Shaib, *Middle East respiratory syndrome coronavirus (MERS-CoV): A review.* Germs, 2019. **9**(1): p. 35-42.
- 8. Vijayanand, P., Wilkins E, Woodhead M, Severe acute respiratory syndrome (SARS): a review. Clin Med, 2004. **4**: p. 152-160.
- 9. Brake, S.J., et al., *Smoking upregulates angiotensin-converting enzyme-2 receptor: a potential adhesion site for novel coronavirus SARS-CoV-2 (Covid-19)*. Journal of Clinical Medicine, 2020. **9**(3): p. 841.
- 10. Hassan, S.A., et al., *Coronavirus (COVID-19): a review of clinical features, diagnosis, and treatment.* 2020. **12**(3): p. e7355.
- 11. Eapen, M.S., et al., *Dysfunctional immunity and microbial adhesion molecules in smoking-induced pneumonia.* 2019. **199**(2): p. 250-251.
- 12. Han, L., et al., *Smoking and influenza-associated morbidity and mortality: a systematic review and meta-analysis.* 2019. **30**(3): p. 405-417.
- 13. Lawrence, H., et al., *Cigarette smoking and the occurrence of influenza–Systematic review.* 2019. **79**(5): p. 401-406.
- 14. Alqahtani, J.S., et al., *Prevalence, severity and mortality associated with COPD and smoking in patients with COVID-19: a rapid systematic review and meta-analysis.* 2020. **15**(5): p. e0233147.
- 15. Guan, W.-j., et al., *Clinical characteristics of coronavirus disease 2019 in China.* 2020. **382**(18): p. 1708-1720.
- 16. Zhang, J.-j., et al., *Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China*. Allergy, 2020. **75**(7): p. 1730-1741.
- 17. Farsalinos, K., et al., *COVID-19 and the nicotinic cholinergic system.* European Respiratory Journal, 2020. **56**(1): p. 2001589.
- 18. Leung, J.M. and D.D. Sin, *Smoking, ACE-2, and COVID-19: Ongoing Controversies*. European Respiratory Journal, 2020: p. 2001759.
- 19. Hoffmann, M., et al., SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell, 2020. **181**(2): p. 271-280.e8.
- 20. Li, F., et al., Structure of SARS coronavirus spike receptor-binding domain complexed with receptor. 2005. **309**(5742): p. 1864-1868.
- 21. Coutard, B., et al., The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. 2020. **176**: p. 104742.
- 22. Heurich, A., et al., *TMPRSS2* and *ADAM17* cleave ACE2 differentially and only proteolysis by TMPRSS2 augments entry driven by the severe acute respiratory syndrome coronavirus spike protein. J Virol, 2014. **88**(2): p. 1293-307.

- 23. Bestle, D., et al., *TMPRSS2* and furin are both essential for proteolytic activation of SARS-CoV-2 in human airway cells. Life Sci Alliance, 2020. **3**(9): p. e202000786.
- 24. Gheblawi, M., et al., *Angiotensin-converting enzyme 2: SARS-CoV-2 receptor and regulator of the renin-angiotensin system: celebrating the 20th anniversary of the discovery of ACE2*. 2020. **126**(10): p. 1456-1474.
- 25. Rey-Parra, G., et al., *Angiotensin converting enzyme 2 abrogates bleomycin-induced lung injury.* 2012. **90**(6): p. 637-647.
- 26. Zheng, Y.-Y., et al., COVID-19 and the cardiovascular system. 2020. 17(5): p. 259-260.
- 27. Hikmet, F., et al., *The protein expression profile of ACE2 in human tissues*. Molecular systems biology, 2020. **16**(7): p. e9610-e9610.
- 28. Wang, K., M. Gheblawi, and G.Y.J.C. Oudit, *Angiotensin converting enzyme 2: a double-edged sword.* 2020.
- 29. Kuba, K., Y. Imai, and J.M.J.C.o.i.p. Penninger, *Angiotensin-converting enzyme 2 in lung diseases*. 2006. **6**(3): p. 271-276.
- 30. Liu, Y., et al., Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. 2020. **63**(3): p. 364-374.
- 31. Lurie, N., et al., Developing Covid-19 vaccines at pandemic speed. 2020. **382**(21): p. 1969-1973.
- 32. Jiménez-Ruiz, C.A., et al., *COVID-19 and Smoking: A Systematic Review and Meta-Analysis of the Evidence.* Arch Bronconeumol, 2021. **57 Suppl 1**: p. 21-34.
- 33. Hagenbjörk-Gustafsson, A., et al., *Determinants of personal exposure to some carcinogenic substances and nitrogen dioxide among the general population in five Swedish cities*. Journal of Exposure Science & Environmental Epidemiology, 2014. **24**(4): p. 437-443.
- 34. Strzelak, A., et al., *Tobacco Smoke Induces and Alters Immune Responses in the Lung Triggering Inflammation, Allergy, Asthma and Other Lung Diseases: A Mechanistic Review.* Int J Environ Res Public Health, 2018. **15**(5).
- 35. van Zyl-Smit, R.N., G. Richards, and F.T. Leone, *Tobacco smoking and COVID-19 infection*. The Lancet Respiratory Medicine, 2020. **8**(7): p. 664-665.
- 36. Nam, H.-S., et al., *High fatality rates and associated factors in two hospital outbreaks of MERS in Daejeon, the Republic of Korea*. International Journal of Infectious Diseases, 2017. **58**: p. 37-42.
- 37. Zhang, J.J., et al., *Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China.* Allergy, 2020. **75**(7): p. 1730-1741.
- 38. Ujjan, I.D., et al., *The clinical and demographical profile of Coronavirus illness: The tale of Tablighi Jamaat and Zaireen in Quarantine / Isolation center at Sukkur and Hyderabad.* Pak J Med Sci, 2020. **36**(Covid19-s4): p. S12-s16.
- 39. Guan, W.J., et al., *Comorbidity and its impact on 1590 patients with COVID-19 in China: a nationwide analysis.* Eur Respir J, 2020. **55**(5).
- 40. Zhou, F., et al., Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet, 2020. **395**(10229): p. 1054-1062.
- 41. Liu, W., et al., Analysis of factors associated with disease outcomes in hospitalized patients with 2019 novel coronavirus disease. Chin Med J (Engl), 2020. **133**(9): p. 1032-1038.
- 42. Chen, X., et al., Retrospective study on the epidemiological characteristics of 139 patients with novel coronavirus pneumonia on the effects of severity. Chongqing Med, 2020: p. 1-9.
- 43. Guan, W.J., et al., *Clinical Characteristics of Coronavirus Disease 2019 in China.* N Engl J Med, 2020. **382**(18): p. 1708-1720.
- 44. Huang, C., et al., *Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China*. Lancet, 2020. **395**(10223): p. 497-506.
- 45. Guan, W.-j., et al., *Clinical Characteristics of Coronavirus Disease 2019 in China*. New England Journal of Medicine, 2020. **382**(18): p. 1708-1720.

- 46. Zhao, Q., et al., *The impact of COPD and smoking history on the severity of COVID-19: A systemic review and meta-analysis.* J Med Virol, 2020. **92**(10): p. 1915-1921.
- 47. Zheng, Z., et al., Risk factors of critical & mortal COVID-19 cases: A systematic literature review and meta-analysis. J Infect, 2020. **81**(2): p. e16-e25.
- 48. Patanavanich, R. and S.A. Glantz, *Smoking Is Associated With COVID-19 Progression: A Meta-analysis*. Nicotine Tob Res, 2020. **22**(9): p. 1653-1656.
- 49. Emami, A., et al., *Prevalence of Underlying Diseases in Hospitalized Patients with COVID-19: a Systematic Review and Meta-Analysis.* Arch Acad Emerg Med, 2020. **8**(1): p. e35.
- 50. Lippi, G. and B.M. Henry, *Active smoking is not associated with severity of coronavirus disease* 2019 (COVID-19). Eur J Intern Med, 2020. **75**: p. 107-108.
- 51. Farsalinos, K., A. Barbouni, and R. Niaura, *Systematic review of the prevalence of current smoking among hospitalized COVID-19 patients in China: could nicotine be a therapeutic option?* Internal and emergency medicine, 2020. **15**(5): p. 845-852.
- 52. Rossato, M., et al., *Current smoking is not associated with COVID-19.* The European respiratory journal, 2020. **55**(6): p. 2001290.
- 53. Smith, J.C., et al., *Cigarette Smoke Exposure and Inflammatory Signaling Increase the Expression of the SARS-CoV-2 Receptor ACE2 in the Respiratory Tract.* Developmental Cell, 2020. **53**(5): p. 514-529.e3.
- 54. Hung, Y.H., et al., Alternative Roles of STAT3 and MAPK Signaling Pathways in the MMPs Activation and Progression of Lung Injury Induced by Cigarette Smoke Exposure in ACE2 Knockout Mice. Int J Biol Sci, 2016. **12**(4): p. 454-65.
- 55. Yilin, Z., N. Yandong, and J.J.B. Faguang, *Role of angiotensin-converting enzyme (ACE) and ACE2 in a rat model of smoke inhalation induced acute respiratory distress syndrome*. 2015. **41**(7): p. 1468-1477.
- 56. Radzikowska, U., et al., *Distribution of ACE2, CD147, CD26, and other SARS-CoV-2 associated molecules in tissues and immune cells in health and in asthma, COPD, obesity, hypertension, and COVID-19 risk factors.* Allergy, 2020. **75**(11): p. 2829-2845.
- 57. Brake, S.J., et al., *Smoking Upregulates Angiotensin-Converting Enzyme-2 Receptor: A Potential Adhesion Site for Novel Coronavirus SARS-CoV-2 (Covid-19)*. J Clin Med, 2020. **9**(3).
- 58. Saheb Sharif-Askari, N., et al., *Airways Expression of SARS-CoV-2 Receptor, ACE2, and TMPRSS2 Is Lower in Children Than Adults and Increases with Smoking and COPD.* Molecular Therapy Methods & Clinical Development, 2020. **18**: p. 1-6.
- 59. Cruz, T., et al., *Multi-level immune response network in mild-moderate Chronic Obstructive Pulmonary Disease (COPD).* Respiratory Research, 2019. **20**(1): p. 152.
- 60. Kim, W.J., et al., *Comprehensive Analysis of Transcriptome Sequencing Data in the Lung Tissues of COPD Subjects.* International Journal of Genomics, 2015. **2015**: p. 206937.
- 61. McDonough, J.E., et al., *Transcriptional regulatory model of fibrosis progression in the human lung.* JCI Insight, 2019. **4**(22): p. e131597.
- 62. Pardo, A., et al., *Up-regulation and profibrotic role of osteopontin in human idiopathic pulmonary fibrosis.* PLoS Med, 2005. **2**(9): p. e251.
- 63. Baumgartner, K.B., et al., *Cigarette smoking: a risk factor for idiopathic pulmonary fibrosis.* Am J Respir Crit Care Med, 1997. **155**(1): p. 242-8.
- 64. Laniado-Laborín, R., Smoking and chronic obstructive pulmonary disease (COPD). Parallel epidemics of the 21 century. International journal of environmental research and public health, 2009. **6**(1): p. 209-224.
- 65. Lippi, G. and B.M. Henry, *Chronic obstructive pulmonary disease is associated with severe coronavirus disease 2019 (COVID-19)*. Respiratory Medicine, 2020. **167**: p. 105941.

- Zhao, Q., et al., *The impact of COPD and smoking history on the severity of COVID-19: A systemic review and meta-analysis.* Journal of Medical Virology, 2020. **92**(10): p. 1915-1921.
- 67. Crouser, E.D., et al., *Gene expression profiling identifies MMP-12 and ADAMDEC1 as potential pathogenic mediators of pulmonary sarcoidosis*. Am J Respir Crit Care Med, 2009. **179**(10): p. 929-38.
- 68. Voraphani, N., et al., *An airway epithelial iNOS-DUOX2-thyroid peroxidase metabolome drives Th1/Th2 nitrative stress in human severe asthma.* Mucosal immunology, 2014. **7**(5): p. 1175-1185.
- 69. Matusiak, M. and C.M. Schürch, *Expression of SARS-CoV-2 entry receptors in the respiratory tract of healthy individuals, smokers and asthmatics*. Respiratory Research, 2020. **21**(1): p. 252.
- 70. Liu, Y., et al., *Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury.* Sci China Life Sci, 2020. **63**(3): p. 364-374.
- 71. Leung, J.M., et al., ACE-2 expression in the small airway epithelia of smokers and COPD patients: implications for COVID-19. Eur Respir J, 2020. **55**(5).
- 72. Ziegler, C.G.K., et al., SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues. Cell, 2020. **181**(5): p. 1016-1035.e19.
- 73. Chappell, G.A., et al., Variation in DNA-Damage Responses to an Inhalational Carcinogen (1,3-Butadiene) in Relation to Strain-Specific Differences in Chromatin Accessibility and Gene Transcription Profiles in C57BL/6J and CAST/EiJ Mice. Environ Health Perspect, 2017. 125(10): p. 107006.
- 74. Citrin, D.E., et al., *Role of type II pneumocyte senescence in radiation-induced lung fibrosis.* J Natl Cancer Inst, 2013. **105**(19): p. 1474-84.
- 75. Haston, C.K., et al., *Strain-dependent pulmonary gene expression profiles of a cystic fibrosis mouse model.* Physiological Genomics, 2006. **25**(2): p. 336-345.
- 76. Kozul, C.D., et al., *Chronic exposure to arsenic in the drinking water alters the expression of immune response genes in mouse lung.* Environmental health perspectives, 2009. **117**(7): p. 1108-1115.
- 77. Cai, G., et al., *Tobacco smoking increases the lung gene expression of ACE2, the receptor of SARS-CoV-2*. Am J Respir Crit Care Med, 2020. **201**(12): p. 1557-1559.
- 78. Lukassen, S., et al., SARS-CoV-2 receptor ACE2 and TMPRSS2 are primarily expressed in bronchial transient secretory cells. The EMBO Journal, 2020. **n/a**(n/a): p. e105114.
- 79. Tomita, K., et al., Coordinate Recruitment of E-Cadherin and ALCAM to Cell–Cell Contacts by  $\alpha$ -Catenin. Biochemical and Biophysical Research Communications, 2000. **267**(3): p. 870-874.
- 80. Farkas, A.E., et al., *HNF4α Regulates Claudin-7 Protein Expression during Intestinal Epithelial Differentiation*. The American Journal of Pathology, 2015. **185**(8): p. 2206-2218.
- 81. González-Mariscal, L., et al., *Tight junction proteins*. Progress in Biophysics and Molecular Biology, 2003. **81**(1): p. 1-44.
- 82. Fahy, J.V. and B.F. Dickey, *Airway Mucus Function and Dysfunction*. New England Journal of Medicine, 2010. **363**(23): p. 2233-2247.
- 83. Rusinova, I., et al., *INTERFEROME v2.0: an updated database of annotated interferon-regulated genes.* Nucleic Acids Research, 2012. **41**(D1): p. D1040-D1046.
- 84. Shapira, S.D., et al., *A Physical and Regulatory Map of Host-Influenza Interactions Reveals Pathways in H1N1 Infection*. Cell, 2009. **139**(7): p. 1255-1267.
- 85. Pedersen, K.B., et al., *Dynamics of ADAM17-Mediated Shedding of ACE2 Applied to Pancreatic Islets of Male db/db Mice.* Endocrinology, 2015. **156**(12): p. 4411-25.

- 86. Chakladar, J., et al., Smoking-Mediated Upregulation of the Androgen Pathway Leads to Increased SARS-CoV-2 Susceptibility. International journal of molecular sciences, 2020. **21**(10): p. 3627.
- 87. Lee, A.C., et al., *Tobacco, but not nicotine and flavor-less electronic cigarettes, induces ACE2 and immune dysregulation.* International journal of molecular sciences, 2020. **21**(15): p. 5513.
- 88. Olds, J.L. and N. Kabbani, *Is nicotine exposure linked to cardiopulmonary vulnerability to COVID-* 19 in the general population? Febs j, 2020. **287**(17): p. 3651-3655.
- 89. Leung, J.M., C.X. Yang, and D.D. Sin, *COVID-19 and nicotine as a mediator of ACE-2*. Eur Respir J, 2020. **55**(6): p. 2001261.
- 90. Russo, P., et al., COVID-19 and smoking: is nicotine the hidden link? Eur Respir J, 2020. 55(6).
- 91. Stefano, G.B., T. Esch, and R.M. Kream, *Potential Immunoregulatory and Antiviral/SARS-CoV-2 Activities of Nitric Oxide.* Med Sci Monit, 2020. **26**: p. e925679.
- 92. Caruso, M., et al., Role of Cigarette Smoke on ACE-2 Protein Membrane Expression in Bronchial Epithelial Cells Using an Air-Liquid Interface Model. Frontiers in Pharmacology, 2020. In Press.
- 93. Stewart, D.L., et al., *Effect of inhaled nitric oxide on oxygen therapy, mechanical ventilation, and hypoxic respiratory failure.* Minerva Pediatr, 2018. **70**(1): p. 51-58.
- 94. Hedenstierna, G., et al., *Nitric oxide dosed in short bursts at high concentrations may protect against Covid 19.* Nitric Oxide, 2020. **103**: p. 1-3.
- 95. Gupte, M., et al., *Angiotensin converting enzyme 2 contributes to sex differences in the development of obesity hypertension in C57BL/6 mice*. Arterioscler Thromb Vasc Biol, 2012. **32**(6): p. 1392-9.
- 96. Liu, J., et al., Sex differences in renal angiotensin converting enzyme 2 (ACE2) activity are 176-oestradiol-dependent and sex chromosome-independent. Biology of sex differences, 2010. **1**(1): p. 6-6.
- 97. Cai, G., Bulk and single-cell transcriptomics identify tobacco-use disparity in lung gene expression of ACE2, the receptor of 2019-nCov. medRxiv, 2020.
- 98. Reckelhoff, J.F., *Gender differences in the regulation of blood pressure.* Hypertension, 2001. **37**(5): p. 1199-208.
- 99. McGuire, B.B., et al., *Gender differences in the renin-angiotensin and nitric oxide systems:* relevance in the normal and diseased kidney. Kidney Blood Press Res, 2007. **30**(2): p. 67-80.
- 100. Rabi, D.M., et al., *Reporting on sex-based analysis in clinical trials of angiotensin-converting enzyme inhibitor and angiotensin receptor blocker efficacy.* The Canadian journal of cardiology, 2008. **24**(6): p. 491-496.
- 101. White, M.C., R. Fleeman, and A.C. Arnold, *Sex differences in the metabolic effects of the reninangiotensin system.* Biol Sex Differ, 2019. **10**(1): p. 31.
- 102. Majdic, G., Could Sex/Gender Differences in ACE2 Expression in the Lungs Contribute to the Large Gender Disparity in the Morbidity and Mortality of Patients Infected With the SARS-CoV-2 Virus? Frontiers in Cellular and Infection Microbiology, 2020. **10**: p. 327.
- 103. The Novel Coronavirus Pneumonia Emergency Response Epidemiology Team, *The Epidemiological Characteristics of an Outbreak of 2019 Novel Coronavirus Diseases (COVID-19) China, 2020[J].* China CDC Weekly, 2020. **2**(8): p. 113-122.
- 104. Fabiani, M., et al., *Epidemiological characteristics of COVID-19 cases in non-Italian nationals notified to the Italian surveillance system.* Eur J Public Health, 2021. **31**(1): p. 37-44.
- 105. Kalidhindi, R.S.R., et al., Sex steroids skew ACE2 expression in human airway: a contributing factor to sex differences in COVID-19? American Journal of Physiology-Lung Cellular and Molecular Physiology, 2020. **319**(5): p. L843-L847.

- 106. Foresta, C., M.S. Rocca, and A. Di Nisio, *Gender susceptibility to COVID-19: a review of the putative role of sex hormones and X chromosome*. Journal of Endocrinological Investigation, 2020: p. 1-6.
- 107. Voinsky, I. and D. Gurwitz, *Smoking and COVID-19: Similar bronchial ACE2 and TMPRSS2* expression and higher TMPRSS4 expression in current versus never smokers. Drug Development Research, 2020. **81**(8): p. 1073-1080.
- 108. Wang, W., et al., *Cigarette smoking has a positive and independent effect on testosterone levels.* Hormones (Athens), 2013. **12**(4): p. 567-77.
- 109. Qing, E., et al., Evaluating MERS-CoV Entry Pathways. Methods Mol Biol, 2020. 2099: p. 9-20.
- 110. Benowitz, N.L., *Drug therapy. Pharmacologic aspects of cigarette smoking and nicotine addiction.* N Engl J Med, 1988. **319**(20): p. 1318-30.
- 111. Seeman, J.I., M. Dixon, and H.J. Haussmann, *Acetaldehyde in mainstream tobacco smoke:* formation and occurrence in smoke and bioavailability in the smoker. Chem Res Toxicol, 2002. **15**(11): p. 1331-50.
- 112. Burns, D.M., Cigarettes and cigarette smoking. Clin Chest Med, 1991. 12(4): p. 631-42.
- 113. Lim, T.K., et al., *Effects of nonsteroidal anti-inflammatory drugs on the bronchial hyperresponsiveness of middle-aged male smokers*. Eur Respir J, 1990. **3**(8): p. 872-9.
- 114. Siew, L.Q.C., et al., Cigarette smoking increases bronchial mucosal IL-17A expression in asthmatics, which acts in concert with environmental aeroallergens to engender neutrophilic inflammation. Clin Exp Allergy, 2017. **47**(6): p. 740-750.
- 115. Son, E.S., et al., *Effects of antioxidants on oxidative stress and inflammatory responses of human bronchial epithelial cells exposed to particulate matter and cigarette smoke extract.* Toxicol In Vitro, 2020. **67**: p. 104883.
- 116. Marie E, K., et al., *Characteristics Associated with Hospitalization Among Patients with COVID-19 Metropolitan Atlanta, Georgia, March–April 2020.* MMWR Morb Mortal Wkly Rep, 2020. **69**(25): p. 790-794.
- 117. Karanasos, A., et al., *Impact of Smoking Status on Disease Severity and Mortality of Hospitalized Patients With COVID-19 Infection: A Systematic Review and Meta-analysis.* Nicotine & Tobacco Research, 2020. **22**(9): p. 1657-1659.
- 118. McAlinden, K.D., et al., *COVID-19 and vaping: risk for increased susceptibility to SARS-CoV-2 infection?* European Respiratory Journal, 2020. **56**(1): p. 2001645.
- 119. Reidel, B., et al., *E-Cigarette Use Causes a Unique Innate Immune Response in the Lung, Involving Increased Neutrophilic Activation and Altered Mucin Secretion.* Am J Respir Crit Care Med, 2018. **197**(4): p. 492-501.
- 120. Sussman, R.A., E. Golberstein, and R. Polosa, *Aerial Transmission of the SARS-CoV-2 Virus through Environmental E-Cigarette Aerosols: Implications for Public Policies.* International journal of environmental research and public health, 2021. **18**(4): p. 1437.
- 121. Gaiha, S.M., J. Cheng, and B. Halpern-Felsher, *Association Between Youth Smoking, Electronic Cigarette Use, and COVID-19.* J Adolesc Health, 2020. **67**(4): p. 519-523.
- 122. Kale, D., et al., Associations between vaping and Covid-19: Cross-sectional findings from the HEBECO study. Drug and Alcohol Dependence, 2021. **221**: p. 108590.
- 123. Caponnetto, P., et al., Smoking behavior and psychological dynamics during COVID-19 social distancing and stay-at-home policies: A survey. Health psychology research, 2020. **8**(1): p. 9124-9124.
- 124. Rolain, J.M., P. Colson, and D. Raoult, *Recycling of chloroquine and its hydroxyl analogue to face bacterial, fungal and viral infections in the 21st century.* Int J Antimicrob Agents, 2007. **30**(4): p. 297-308.

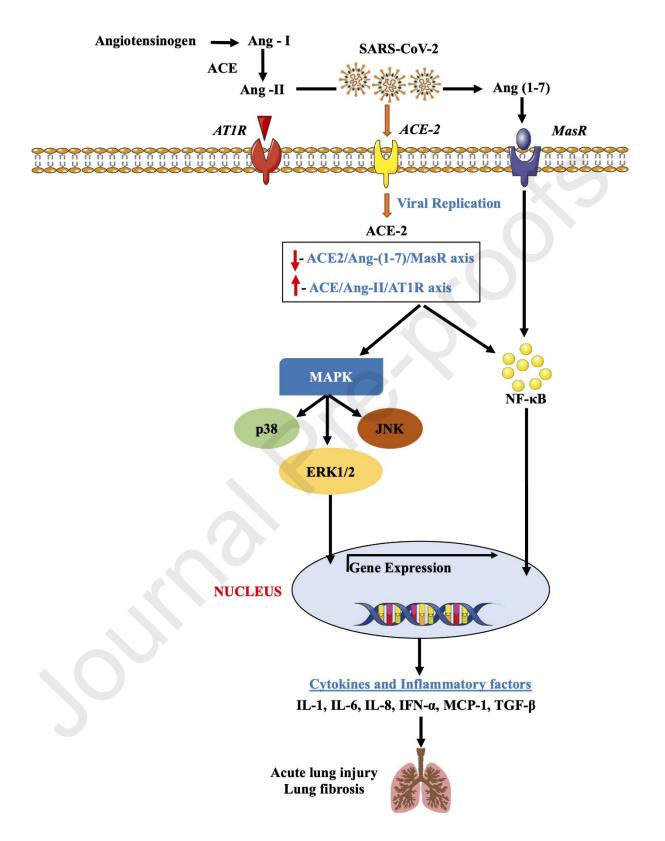
- 125. Dong, L., S. Hu, and J. Gao, *Discovering drugs to treat coronavirus disease 2019 (COVID-19)*. Drug Discoveries & Therapeutics, 2020. **14**(1): p. 58-60.
- 126. Savarino, A., et al., *New insights into the antiviral effects of chloroquine.* The Lancet Infectious Diseases, 2006. **6**(2): p. 67-69.
- 127. Fragkou, P.C., et al., *Review of trials currently testing treatment and prevention of COVID-19*. Clinical Microbiology and Infection, 2020. **26**(8): p. 988-998.
- 128. Sanders, J.M., et al., *Pharmacologic treatments for coronavirus disease 2019 (COVID-19): a review.* 2020. **323**(18): p. 1824-1836.
- 129. Wang, M., et al., *Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro*. Cell Research, 2020. **30**(3): p. 269-271.
- 130. Zhou, Y., et al., *Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2.* Cell Discovery, 2020. **6**(1): p. 14.
- 131. Lin, S.M., et al., Structure-Based Stabilization of Non-native Protein-Protein Interactions of Coronavirus Nucleocapsid Proteins in Antiviral Drug Design. J Med Chem, 2020. **63**(6): p. 3131-3141.
- 132. Gordon, D.E., et al., *A SARS-CoV-2 protein interaction map reveals targets for drug repurposing.* Nature, 2020. **583**(7816): p. 459-468.
- 133. Kandeel, M. and M. Al-Nazawi, *Virtual screening and repurposing of FDA approved drugs against COVID-19 main protease.* Life Sci, 2020. **251**: p. 117627.
- 134. Yamaya, M., et al., The serine protease inhibitor camostat inhibits influenza virus replication and cytokine production in primary cultures of human tracheal epithelial cells. Pulmonary pharmacology & therapeutics, 2015. **33**: p. 66-74.
- 135. Uno, Y., *Camostat mesilate therapy for COVID-19.* Internal and emergency medicine, 2020: p. 1-2.
- 136. Bertram, J.F. and A.W. Rogers, *Recovery of bronchial epithelium on stopping smoking*. British medical journal (Clinical research ed.), 1981. **283**(6306): p. 1567-1569.
- 137. Lee, J.J., et al., Long-term impact of smoking on lung epithelial proliferation in current and former smokers. J Natl Cancer Inst, 2001. **93**(14): p. 1081-8.
- 138. Lutchman, D., Could the smoking gun in the fight against Covid-19 be the (rh) ACE2? European Respiratory Journal, 2020: p. 2001560.
- 139. Monteil, V., et al., *Inhibition of SARS-CoV-2 Infections in Engineered Human Tissues Using Clinical-Grade Soluble Human ACE2*. Cell, 2020. **181**(4): p. 905-913.e7.
- 140. Bundgaard, H. Recombinant Human Angiotensin-converting Enzyme 2 (rhACE2) as a Treatment for Patients With COVID-19 (APN01-COVID-19). 2020 October 6, 2020 [cited 2020 October 24]; Available from: https://clinicaltrials.gov/ct2/show/NCT04335136.
- 141. Chen, F., et al., *In vitro susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds.* Journal of Clinical Virology, 2004. **31**(1): p. 69-75.
- 142. Sheahan, T.P., et al., *Comparative therapeutic efficacy of remdesivir and combination lopinavir, ritonavir, and interferon beta against MERS-CoV.* Nature Communications, 2020. **11**(1): p. 222.
- 143. Chen, J.L., Y.; Xi, X.; Liu, P.; Li, F.; Li, T, *Efficacies of lopinavir/ritonavir and abidol in the treatment of novel coronavirus pneumonia*. Chin. J. Infect, 2020. **38**: p. E008.
- 144. Verdugo-Paiva, F., et al., *Lopinavir-ritonavir for COVID-19: A living systematic review*. Medwave, 2020. **20**(6): p. e7967.
- 145. Wu, A., et al., Genome Composition and Divergence of the Novel Coronavirus (2019-nCoV)
  Originating in China. Cell Host & Microbe, 2020. **27**(3): p. 325-328.
- 146. Wu, C., et al., *Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods.* Acta Pharm Sin B, 2020. **10**(5): p. 766-788.

infected pneumonia in Wuhan, China. 2020. **323**(11): p. 1061-1069.

147.

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Author Contributions Statement
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Declaration of interests
oxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.
☐The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:



Highlights

- Summary of correlation between smoking and the expression of SARS-CoV-2 key entry genes, clinical manifestations, and disease progression
- Smoking-related cardiovascular and respiratory pre-conditions are anticipated risk factors for COVID-19
- The impact of smoking on ACE-2 and TMPRSS2 receptors expression is still controversial